

Antioxidant Activity Comparison between Young and Old Malaka Fruit (*Phyllanthus emblica* L.) Extracts from Bandung, Indonesia

Sani Fitriansyah N. *, Sartika Amalia and Rival Ferdiansyah

Indonesia School of Pharmacy, Jl. Soekarno-Hatta, No.354, Bandung, INDONESIA

*saninurlaela@stfi.ac.id

Abstract

Phyllanthus emblica L. is a native plant from Indonesia which has been shown to have biological activity as antioxidant. The maturity of a plant could affect the different types and quantities of secondary metabolites and cause differences of biological activities. The objective of this research was to compare the antioxidant activity of young and old *P. emblica* fruit extracts from Bandung-Indonesia. Extraction was performed using macerator with different polarity of solvents as follow: n-hexane, ethyl acetate and ethanol. The extracts were vaporated using rotavapor. The antioxidant activity was tested using DPPH assay and monitored by TLC in order to find out the existing compounds which were responsible for antioxidant activity.

The results showed that the extracts from *P. emblica* performed antioxidant activity with IC_{50} values of ≤ 50 $\mu\text{g/mL}$. Therefore, they were classified as very strong antioxidant. IC_{50} value of young *P. emblica* fruit extract showed the smaller value than IC_{50} value of old *P. emblica* fruit extract. The strongest antioxidant activity of all *P. emblica* extracts was showed by ethyl acetate extract from young *P. emblica* and it was shown by phenol substances. As conclusion, the young *P. emblica* fruit extract had stronger antioxidant activity than the old *P. emblica* fruit extract.

Keywords: Young and old fruit, *Phyllanthus emblica* L., antioxidant, DPPH assay.

Introduction

In the world of pharmacy, medicinal plants are used as raw material for medicine. Many of the researches provide biological activities information from various plants. Biological activity of a plant could be due to the presence of secondary metabolites in that medicinal plant¹. Secondary metabolites in medicinal plant such as phenols and flavonoids are often referred to as bioactive components for various biological activities².

Flavonoids were widely used for some biological activities such as antioxidant³. Different types of phenol compounds could cause differences of antioxidant potency and antibacterial strength⁴.

Type and quantity of secondary metabolite in medicinal plant could cause differences in the strenght and variety of biology activity potency¹. Physiological processes in a plant, the maturity part of plant and environmental conditions⁵ such as sunlight condition, air pressure and temperature could be factors to different types and quantity secondary metabolites^{6,7}.

Phyllanthus emblica L. is one of the main species of plants that grow in various countries including Indonesia (West Java). *Phyllanthus emblica* L. has a variety of biological activities such as antibacterial against *Escherichia coli*, *Pseudomonas aeruginosa*, *Staphylococcus aureus* and *Bacillus subtilis*⁸; antifungal against *Aspergillus Niger*, *Candida albicans*, *Penicillium nasturtium*⁹; inflammatory¹⁰; antidiarrheal¹¹⁻¹³; antioxidant caused by phenolic groups¹² and antioxidants for anticancer¹⁴. However, the biology activity for antioxidants from maturity difference of *Phyllanthus emblica* L. fruit had not been reported. According to Hasan et al¹⁵, tannin and phenolic group were seconday metabolites in *P. emblica* fruit and could be responsible for antioxidant activity.

Material and Methods

Materials: DPPH (2,2-diphenyl-1-picrylhydrazyl) and ascorbic acid were purchased from Sigma-Aldrich (MO, USA), young and old fruit of *P. emblica*, methanol P.a, ethanol, ethyl acetate, n-hexane and others analytical materials were used in this study.

Methods

Sample preparation: *P. emblica* of young and old fruit were freshly collected from Bale Endah-Bandung, West Java-Indonesia on December 2016. *P. emblica* fruits were sorted, washed, dried at 40°C - 45°C and grinded into powder form.

Extraction: *P. emblica* of young and old fruit were extracted by using maserator with different polarity of solvent which were n-hexane, ethyl acetate and ethanol. Each sample was extracted with n-hexane in three repetitions, then the residue was extracted with ethyl acetat in three times repetition and the end residue was extracted with ethanol in three times repetition. Each extract was concentrated using rotary vaporator and resulted into thick n-hexane extract of young fruit *P. emblica* (YN), ethyl acetate extract (YE) and ethanol extract (YL), thick n-hexane extract of old fruit *P. emblica* (ON), ethyl acetate extracts (OE) and ethanol extract (OL).

Monitoring of secondary metabolites: Monitoring of secondary metabolites was performed against simplicia and fruit extract of *P.emblica* (YH, YE, YL, OH, OE and OL). A phenolic compound was identified by using FeCl_3 10% reagent, a flavonoid using the amyl alcohol reagent, a tannin using gelatin, an alkaloid using Dragendorf and Mayer reagents, quinones using KOH 5%, saponins showing with a constant foam \pm 10 minutes in water extracts, monoterpen and sesquiterpen using 10% solution of vanillin in H_2SO_4 , steroid and triterpenoid using Lieberman-Burchard reagent¹⁶.

Antioxidant activity using the Blois method: Antioxidant activity was adopted from the Blois methode¹⁷. DPPH solution of 50 ppm was used as control and ascorbic acid was used as antioxidant standard. Sample was prepared in various concentrations, then added with 50 ppm of DPPH (volume 1:1) and then incubated for 30 minutes. After incubation for 30 minutes, the absorbance was measured at λ 517 nm using a UV-visible spectrophotometry. Methanol was used as blank. Antioxidant activity was measured as a percentage of the sample against DPPH decrease absorbance. IC_{50} of scavenge to DPPH was determined using the calibration curve of the antioxidant activity of samples in various concentrations.

Monitoring of antioxidant compound in extract using TLC: Monitoring of antioxidant compound was performed against extract which had the smallest value of IC_{50} . Admixture of ethyl acetate and n-hexane were used as eluents. FeCl_3 1% was used for detecting the existence of phenolic compounds and citroboric 1% was used for detecting the existence of flavonoid compounds.

Results and Discussion

Extraction: Each sample had differences of extract content per gram of simplicia and organoleptic by colors. Extract content per gram of *P.emblica* fruit simplisia was shown in fig. 1. Extract content of YE was the highest. It meant that ethyl acetat was the best solvent between n-hexane and ethanol for getting extract content of young *P.emblica* fruit. The organoleptic result showed green color for YN and ON, brown to dark for YE and OE and dark red for YL and OL.

Monitoring of secondary metabolite: A secondary metabolite of YL and OL is differences by tannin. Chemical compound of *P. Emblica* extract had been widely reported including tannin and phenol compounds. The production of secondary metabolites could be affected by the maturity of the plant¹⁸. The results showed that young and old fruits of *P. emblica* had some chemical compounds of flavonoid, phenolic, quinone and saponin. The differences in chemical compounds were indicated by the presence of tannin in young fruit of *P.emblica* but not in old fruit of *P.emblica* like that shown by YL.

Antioxidant activity and IC_{50} to scavenging of DPPH: Antioxidant activity of a sample is shown by percentage of sample in 50 ppm to scavenging of DPPH. The antioxidant activity test was performed using Blois¹⁷ method with DPPH. DPPH is a relatively stable free radical¹⁹. The DPPH solution is purple and shows absorption at 515-520 nm wavelength²⁰. In case of scavenging of DPPH from the sample it will show a color change²¹. Antioxidant activity of fruit extract *P.emblica* from Bandung had differences which showed in range 54-94% for young fruit extract of *P.emblica* and 52-93% for old fruit extract of *P.emblica*.

Antioxidant activity of young fruit extract of *P.emblica* from Bandung was higher than old fruit extract of *P.emblica*. YE was the highest antioxidant activity when compared to YN, YL, ON, OE and OL. This result suggested that antioxidant compound in YE had the higher potency to scavenge the free radical DPPH. The previous reserach by Sumalatha²² showed that antioxidant activity with scavenging of DPPH on ethanol extract fruit of *P.emblica* was 71,75%, water extract fruit of *P.emblica* showed IC_{50} value to scavenging of DPPH as 51,3 $\mu\text{g/mL}$ ²³ and percentage of scavenging free radical DPPH at 100 μg concentration of extract leaves *P.emblica* was 82.053%²⁴. The maturity part of plant could influence the type and quantity of secondary metabolite¹. The type and quantity of secondary metabolite could influence the biology activity of medicinal plant.

IC_{50} to scavenging of DPPH from each extract could be seen in fig. 3. IC_{50} to scavenging of DPPH from young fruit extract of *P.emblica* had the smallest value than the old one. It means that a young fruit extract of *P.emblica* had the highest antioxidant activity than the old fruit extract. As we know that IC_{50} is the concentration of sample which can reduce DPPH by 50%. IC_{50} of all sample was compared with the standard of ascorbic acid. The smallest of IC_{50} showed the highest antioxidant activity.

Antioxidant activity can be classified when showed by IC_{50} value. According to Blois¹⁷, sample which had IC_{50} lower than 50 $\mu\text{g/mL}$ was a very strong antioxidant. IC_{50} value is the concentration of samples that can scavenge 50% of free radical DPPH activity. The highest antioxidant activity was indicated by the lowest value of IC_{50} . IC_{50} of DPPH scavenging capacities of YN, YE, YL, ON, OE and OL were compared to IC_{50} of ascorbic acid as a standard. As a result, YE had the lowest value of IC_{50} compared to YN, YL, ON, OE and OL. ON had the highest value of IC_{50} compared to YN, YE, YO, OE and OL. All extracts had a value of IC_{50} lower than 50 $\mu\text{g/mL}$, while IC_{50} DPPH of ascorbic acid was 2.468 $\mu\text{g/mL}$. This result suggested that all fruit extracts of *P.emblica* from Bale Endah-Bandung could be classified as a very strong potential antioxidant.

Monitoring of antioxidant compounds: Monitoring of antioxidant compound was performed on selected extract i.e. YE. It is based on the lowest IC_{50} value which was

compared to other extract. Spot of YE which was positive by spraying of 0.2% DPPH was the same spot which was positive on phenolic compound. It was shown in fig. 4.

Antioxidant activity of extract can be present in suspected compounds that are capable in donating hydrogen on free radicals. Hydrogen donors could be groups of phenolic, flavonoid and carotenoid.^{3,25} Phenolic and flavonoid compounds had the biggest group as compounds which have antioxidant activity in the medicinal plants²⁶. Phenolic and flavonoid compounds were very soluble in polar solvents. Acidic compound of cinamic acid and benzoic acid contributed to the antioxidant activity. Cinnamic acid had the higher antioxidant activity than benzoic acid^{27,28}. Flavonoid compound in extract had a role to antioxidant activity. Position of hydroxyl group at C-3 and ortho position at C-3' and C-4' would increase the antioxidant activity in extract^{28,29} and also with double bonds between C-2 and C-3^{28,29}.

Beside phenolic and flavonoid compound, carotenoid compound in extract was also determined in improving antioxidant activity. Conjugated double bond in carotenoid compounds affected the antioxidant activity^{30,31}. Astaxanthin, β -carotene dan α -tocoferol were determined in improving antioxidant activity. B-carotene was effective

as antioxidant activity by interfering the chain reaction of free radical^{30,32}. Carotenoid compound was very soluble in nonpolar solvent as n-hexane.

TLC was used in monitoring antioxidant compound in extract. YE was monitored with TLC using ethyl acetate-n-hexane as eluent. The spots were identified by FeCl_3 1% for analyzing the presence of phenolic compounds, Citroborat 1% for analyzing the flavonoid compounds and DPPH 0,2% for analyzing the antioxidant compounds. The positive result from FeCl_3 1% would give a brown to black colors in visible on the spotted, Citroborat 1% would give light blue color in spotted by UV lamp at 366 nm and DPPH 0,2% would give a yellow color in spotted with purple background by visible. The results showed that YE performed positive to FeCl_3 1% and Citroborat 1%. The positive spotted by DPPH 0,2% showed the same positive spotted by FeCl_3 1%.

This results suggested that the antioxidant compound were playing role in YE shown by phenolic group. Flavonoid in YE could contribute as antioxidant compound, but not as dominant compound. It was because that flavonoid in YE does not have hydroxyl ortho in C-3' and C-4' and does not have double bond between C-2 and C-3 and does not have hydroxyl position in C-4.

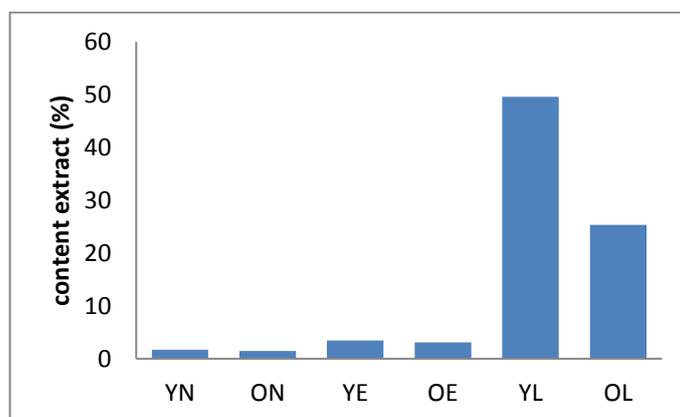


Fig. 1: Content extract per gram of simplicia

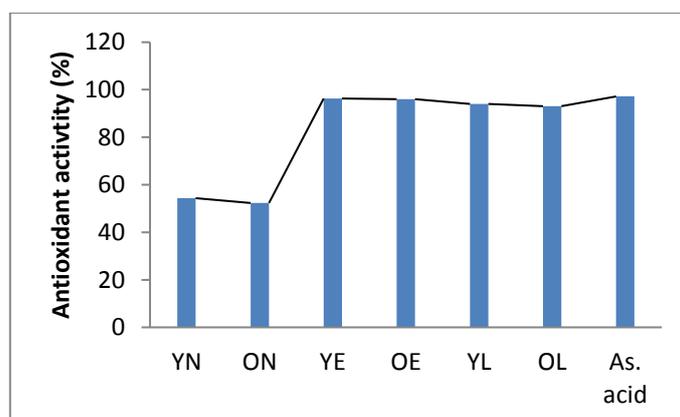


Fig. 2: Antioxidant activity at 50 ppm extracts

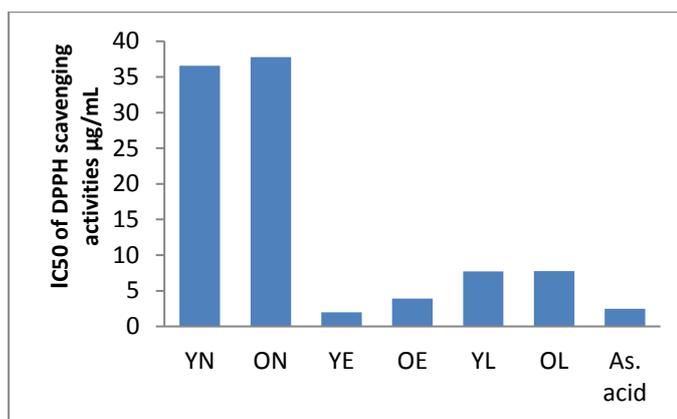


Fig. 3: IC₅₀ of DPPH scavenging activities of extracts

Note : YN (n-hexane extract of young fruit *P.emblica*; YE (ethyl acetate extract); YL (ethanol extract of young fruit *P.emblica*; ON (n-hexane extract of old fruit *P.emblica*; OE (ethyl acetate extracts; OL (ethanol extract of old fruit extract fruit *P.emblica*)

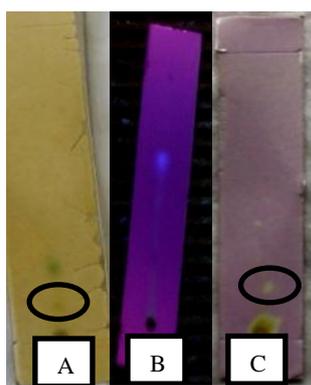


Fig. 4: Antioxidant compound in YE with TLC

Note : A (TLC with FeCl₃ 1%); B (TLC with Citroborat 1%); C (TLC with DPPH 0,2%)

Conclusion

Antioxidant activity of young and old fruit extract of *P.emblica* had differences. Antioxidant activity from ethyl acetate young fruit extract of *P.emblica* had the highest activity than the other extracts. Phenolic compounds caused antioxidant activity of ethyl acetate young fruit extract of *P.emblica*.

References

- Ghasemzadeh A., Jaafar H.Z., Ashkani S., Rahmat A., Juraimi A.S., Puteh A. and Mohamed T.M., Variation in secondary metabolite production as well as antioxidant and antibacterial activities of *Zingiber zerumbet* (L.) at different stages of growth, *BMC Complementary and Alternative Medicine*, **16**, 104 (2016)
- Ghasemzadeh A., Nasiri A., Jaafar H.Z., Baghdadi A. and Ahmad I., Changes in phytochemical synthesis, halcone synthase activity and pharmaceutical qualities of Sababh snake grass (*Clinacanthus nutans* L.) in relationship to plant age, *Molecules*, **19**(11), 32-48 (2014)
- Fidrianny I., Aristya T. and Hartati R., Antioxidant capacities of various leaves extracts from three species of legumes and correlation with total flavonoid, phenolic, carotenoid content, *Int J Pharmacognosy Phytochem Res.*, **7**(3), 628-634 (2015)
- Ghasemzadeh A., Jaafar H.Z. and Rahmat A., Identification and concentration of some flavonoid component in Malaysian young ginger (*Zingiber officinale* Roscoe) varieties by a high-performance liquid chromatography method, *Molecule*, **15**(9), 6231-43 (2010)
- Dai J. and Mumper R.J., Plant phenolic extracton, analysis and their antioxidant and anticancer properties, *Molecule*, **15**(10), 7313-52 (2015)
- Wang S.Y., Bunce J.A. and Mass J., Elevated carbon dioxide increases content of antioxidant compounds in field-grown strawberries, *J Agric Food Chem.*, **51**(15), 4315-4320 (2003)
- Duma Y., Dadomo M., Di Lucca G. and Grolier P., Effects of environmental factors and agricultural techniques on antioxidant content of toamoes, *J Sci Food Agric.*, **83**(5), 369-382 (2003)
- Dhale D.A. and Mogle U.P., Phytochemical screening and antibacterial activity of *Phyllanthus emblica* (L.), *Science Research Reporter*, **1**(3), 138-142 (2011)
- Malliga N.E., Dhanarajan M.S. and Elangovan I., Evaluation of antibacterial and antifungal activity of *Phyllanthus emblica* leaf extract, *Int. J. Pharm. Biol. Sci.*, **2**(2), 59- 66 (2015)
- Krisnaveni M. and Mirunalini S., Therapeutic potential of *Phyllanthus emblica* (amla): the ayurvedic wonder, *J Basic Clin Physiol Pharmacol.*, **21**(1), 93-105 (2010)

11. Treadway L., Amla: Traditional food and medicine, *The Journal of the American Botanical Council*, **31**, 26-42 (1994)
12. Nain P., Saini V. and Sharma S., In-vitro antibacterial and antioxidant activity of *Embllica officinalis* leave extract, *Int J Pharm Pharm Sci.*, **4(1)**, 385-389 (2014)
13. Morton J.F., The emblica (*Phyllantus emblica* L.), *Economy Botany*, **14**, 119-128 (1960)
14. Zhao T., Sun Q., Marques M. and Witcher M., Anticancer of *Phyllantus emblica* (Indian Gooseberry), *Oxidative Medicine and Cellular Logevity*, <http://dx.doi.org/10.1155/2015/950890> (2015)
15. Hasan R., Islam N. and Islam R., Phytochemistry, pharmacological activities and traditional uses of *Embllica officinalis*: A review, *Int. Curr. Pharm. J.*, **5(2)**, 14-21 (2016)
16. Marlina S.D., Suryanti V. and Suyono, Skrining fitokimia dan analisis kromatografi lapis tipis komponen kimia buah labu siam (*Sechium edule* Jacq. Swartz.) dalam ekstrak etanol, *Biofarmasi*, **3(1)**, 26-31 (2005)
17. Blois M.S., Antioxidant determination by the use of stable free radical, *Nature*, **181**, 1199-2000 (1958)
18. Bourgaud F., Gravot A., Milesi S. and Gontier E., Production of plant secondary metabolites: a historical perspective, *Plant Science*, **16**, 839-851 (2001)
19. Sebei K., Gnoum A., Herchi W., Sakouhi F. and Boukhchina S., Lipids, protein, phenolic composition, antioxidant and antibacterial activities of seed of peanut (*Arachis hypogaea* L) cultivated in Tunisia, *Biol Res.*, **46(3)**, 257-263 (2013)
20. Li X.C., Wang X.Z., Chen D.F. and Chen S.Z., Antioxidant activity and mechanism of protochatechuic acid in vitro, *J Funct Food Health Dis.*, **1**, 232-244 (2011)
21. Apak R. et al, Comparative evaluation of various total antioxidant capacity assay applied to phenolic compounds wwith CUPRAC assay, *Molecules*, **1**, 1496-1547 (2007)
22. Sumalatha D., Antioxidant and antitumor activity of *Phyllantus emblica*, *Int. J. Curr. Microbiol. App. Sci.*, **2(5)**, 189-195 (2013)
23. Charoenteeraboon J., Ngamkitidechakul C., Soonthorncharenon N., Jaijoy K. and Sireeratawong S., Antioxidant activities of the standarized water extract from fruit of *Phyllantus emblica* Linn., *J. Sci. Technol.*, **32(6)**, 599-604 (2010)
24. Durga D.M. and Banu N., Study of antioxidant activity of *Phyllantus emblica* L. From chlorophyllin, *Indian J Appl Res.*, **5(3)**, 22-23 (2015)
25. Sharma Neeraj Kant, Sangh Partap, Priyanka Priyanka, Jha Keshari K., Singh Hemant K. and Shrivastava Anil K., Free radical scavenging activity of methanolic extract of *Luffa cylindrica* leaves, *Int J Green Pharm.*, **6(3)**, 231-236 (2012)
26. Fidrianny I., Puspitasari N. and Singgih M., Antioxidant activities, total flavonoid, phenolic, carotenoid of various shells extract from four species of legumes, *Asian J Pharm Sci.*, **7(4)**, 42-46 (2014)
27. Fidrianny I., Darmawati A. and Sukrasno, Antioxidant capacities from different polarities extracts of Cucurbitaceae leaves using FRAP, DPPH assay and correlation with phenolic, flavonoid, carotenoid content, *Int J Pharm Pharm Sci.*, **6(2)**, 58-862 (2014)
28. Heim K.E., Tagliaferro A.R. and Bobilya D.J., Flavonoid antioxidants: chemistry, metabolism and structure-activity relationship, *J Nutr Biochem.*, **13**, 572-584 (2002)
29. Adekunle A.S., Aline A.B., Afolabi O.K. and Rocha J.B.T., Determination of free phenolic, flavonoid contents and antioxidant capacity of ethanolic extracts obtained from leaves of mistletoe (*Tapinanthus globiferus*), *Asian J Pharm Clin Res.*, **5(3)**, 36-41 (2012)
30. Gramza-Michalowska A. and Stachowiak B., The antioxidant potential of carotenoid extract from *Phaffia rhodozyma*, *Acta Sci. Pol., Technol. Aliment.*, **9(2)**, 171-188 (2010)
31. Britton G., Liaaen-Jensen S. and Pfander H., Carotenoids, Handbook, Birkhauser Verlag, Basel, Switzerland (2004)
32. Fiedor J. and Burda K., Potential role of carotenoids as antioxidant in human health and disease, *Nutrients*, **6**, 466-488 (2014).